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Discordant Pleistocene population size histories in a guild of hymenopteran parasitoids

William Walton | Graham N. Stone  | Konrad Lohse 

Institute of Evolutionary Biology,
University of Edinburgh, Edinburgh, UK

Correspondence

Konrad Lohse Institute of Evolutionary
Biology, University of Edinburgh,
Edinburgh EH9 3FL, UK.
Email: konrad.lohse@ed.ac.uk

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Abstract

Signatures of past changes in population size have been detected in genome-wide variation in many species. However, the causes of such demographic changes and the extent to which they are shared across co-distributed species remain poorly understood. During Pleistocene glacial maxima, many temperate European species were confined to southern refugia. While vicariance and range expansion processes associated with glacial cycles have been widely documented, it is unclear whether refugial populations of co-distributed species have experienced shared histories of population size change. We analyse whole-genome sequence data to reconstruct and compare demographic histories during the Quaternary for Iberian refuge populations in a single ecological guild (seven species of chalcid parasitoid wasps associated with oak cynipid galls). For four of these species, we find support for large changes in effective population size (N_e) through the Pleistocene that coincide with major climate events. However, there is little evidence that the timing, direction and magnitude of demographic change are shared across species, suggesting that demographic histories in this guild are largely idiosyncratic, even at the scale of a single glacial refugium.

KEYWORDS

chalcid parasitoids, comparative phylogeography, glacial refugia, population genomics, population size change

1 | INTRODUCTION

Natural populations change in size and range in response to biotic and abiotic factors over both ecological and evolutionary timescales. Given that sequence variation is the only source of information about the evolutionary past of many taxa, there is considerable interest in using genetic data to reconstruct the demographic history of populations (see Beichman et al., 2018, for a recent review). A key question is to what extent past demographic changes have been shaped by climatic events and whether such responses are concordant across co-distributed taxa, or vary with species traits

(Hickerson et al., 2010; Papadopoulou & Knowles, 2016; Stone et al., 2012).

A major aim of phylogeographic studies has been to understand the origins and determinants of spatial structure in natural populations (Avice et al., 1987; Hickerson et al., 2010). Systematic comparisons of demographic histories across co-distributed taxa offer the chance to identify shared abiotic triggers of demographic events (Bermingham & Avice, 1986; Hewitt, 2000; Hickerson et al., 2010). If divergence and admixture between populations are largely the outcome of shared climatic and geological history, one would expect co-distributed and ecologically similar species to show concordant

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histories. Concordance in vicariance times has indeed been found by a number of comparative phylogeographic studies (e.g. Leaché et al., 2007; Stone et al., 2012; Wan et al., 2021; Xue & Hickerson, 2020). However, most species are likely to have undergone complex range shifts and, given that range changes are likely to depend on multiple traits, the idea of strict temporal concordance across species seems implausibly simplistic biologically (Papadopoulou & Knowles, 2016).

For example, many temperate plant and animal taxa (including humans) have colonized the Western Palaearctic in a series of westwards range expansions. Overlaid onto this older longitudinal colonization history is a cyclical history of latitudinal range changes (Hewitt, 2000; Hofreiter & Stewart, 2009). Starting with the onset of major glacial cycles in the mid Pleistocene, the ranges of temperate taxa in Europe were restricted to southern refugia during glacial maxima, primarily in Iberia, Italy and the Balkans (Felinier, 2011; Hewitt, 2000; Hofreiter & Stewart, 2009) and expanded northwards during interglacial periods. As a consequence, modern refugial populations commonly harbour higher genetic diversity than northern populations founded by postglacial range expansion (e.g. Hewitt, 2000; Rokas et al., 2003; Tison et al., 2014; Viales et al., 2016) (but see Comps et al., 2001). While the structuring of genetic diversity by refugia is ubiquitous (Hewitt, 1996), recent comparative studies of the population and speciation history of European taxa have, perhaps unsurprisingly, shown no support for temporal concordance (Bunnefeld et al., 2018; Ebdon et al., 2021).

Bunnefeld et al. (2018) used whole-genome sequence data to compare the timing of divergence and admixture events between three southern refugial populations across 13 co-distributed species in a tritrophic Western Palaearctic community comprising oak (*Quercus*) host plants, cynipid gall wasp herbivores and chalcid parasitoid natural enemies (Stone et al., 2002). The latter are obligate specialists of cynipid galls, allowing the guild of parasitoids to be considered in ecological isolation. Many of the component species show broad longitudinal distributions, extending from Iberia in the West to Iran in the East, and all species studied to date are genetically structured into major southern refuge populations (Lohse et al., 2010, 2012; Nicholls et al., 2010, 2012; Petit et al., 2002; Stone et al., 2012, 2017). Bunnefeld et al. (2018) showed that seven out of

the 13 community members studied colonized refugia in the Balkans and Iberia through westwards range expansion from an eastern origin one or more glacial cycles in the past. However, four species had a discordant, Western origin and two showed no clear signal of a longitudinal history. Bunnefeld et al. (2018) also found little evidence for temporal concordance: although divergence and admixture between refugial populations are broadly concentrated in the late Pleistocene, the timing of divergence and admixture between refugia is to a large extent idiosyncratic across species. However, in order to explore a plausible space of necessarily complex demographic scenarios which capture aspects of both the longitudinal colonization history (divergence between refugia) and the more recent history of latitudinal range shifts (admixture between refugia), Bunnefeld et al. (2018) had to make drastic simplifying assumptions about the second major axis of demographic history: population size change. Specifically, their analyses assumed a single fixed effective population size per refugium. This immediately raises the question of whether the lack of concordance of species' histories found by Bunnefeld et al. (2018) is unique to the broad spatial and deep temporal scale captured by their models, or a more general property of the history of ecological communities which holds over a range of spatio-temporal scales. There are several reasons to expect the degree of temporal concordance to depend on spatial and temporal scales (Papadopoulou & Knowles, 2016). In particular, colonization and admixture histories may be more temporally discordant than population size changes, simply because the former depend on long-distance dispersal events which are both a function of a species' dispersal ability and inherently random (Smith et al., 2014).

Here, we use whole-genome sequence (WGS) data to quantify and compare the histories of population size change within a single glacial refugium across seven chalcid parasitoid species in the oak gall wasp community previously studied by Bunnefeld et al. (2018). We focus on the Iberian refugium, which for most species in the oak gall wasp parasitoid community—including four out of the seven parasitoids studied here (Table 1)—represents the end point of a longitudinal expansion history: Iberian populations have lower genetic diversity than more eastern refugia and show little evidence of population structure (Nicholls et al., 2010; Rokas et al., 2003; Stone et al.,

TABLE 1 Maximum-composite likelihood estimates (MCLE) of parameters under the single-step-change model for Iberian populations of seven species of chalcid parasitoid wasps under a model of a single-step change. Estimates for the time of N_e change (T) are given in thousands of years ago (kya). 95% CIs are shown in brackets. The maximum N_e is scaled relative to π as $Max[N_e]/N_{e,\pi}$. The second column gives the origin of the longitudinal expansion history as inferred by Bunnefeld et al. (2018)

Species	Origin	π	Current $N_e \times 10^3$	Ancestral $N_e \times 10^3$	T (kya)	Relative $Max[N_e]$
<i>Torymus auratus</i>	n/a	0.00509	1,142 (1,126–1,157)	422 (418–425)	100 (94–106)	2.14
<i>Megastigmus stigmatizans</i>	West	0.00054	18 (14–21)	61 (60–61)	10 (9.5–10.4)	1.55
<i>Megastigmus dorsalis</i>	East	0.00142	67 (59–75)	144 (143–145)	12.5 (12.0–13.0)	1.40
<i>Ormyrus nitidulus</i>	East	0.00126	47 (45–49)	194 (192–197)	33 (31–35)	2.13
<i>Ormyrus pomaceus</i>	West	0.00230	207 (207–208)	n/a	n/a	n/a
<i>Eurytoma brunniventris</i>	East	0.00569	523 (521–525)	n/a	n/a	n/a
<i>Cecidostiba fungosa</i>	East	0.00258	234 (233–235)	n/a	n/a	n/a

2007) (for an exception, see Rokas et al. (2001)). Crucially, Bunnefeld et al. (2018) show that Iberian populations are not affected by post-colonization gene flow from other refugia. This allows for a meaningful comparison of Iberian population size histories in isolation from other refugia. Our sampling targets males (five from Iberia, one from the Balkans in each species), whose haploid genome facilitates analysis (Bunnefeld et al., 2018; Hearn et al., 2014). WGS data allow for powerful demographic inference even for non-model organisms. However, given fragmented reference genomes and limited sample sizes available for these species, inference methods must be chosen carefully and, ideally, should use linkage as well as allele frequency information. In particular, inference based only on the site frequency spectrum (SFS) (e.g. Gutenkunst et al., 2009) requires larger samples for accurate inference of recent population history.

We investigate the history of population size change in Iberia using two approaches that use the signal contained in genome-wide variation of a small sample of individuals in different ways (Bunnefeld et al., 2018; Li & Durbin, 2011; Lohse et al., 2011). We use a parametric maximum-composite likelihood (MCL) method based on a blockwise summary of sequence variation (Lohse et al., 2011) to fit a model of a single instantaneous step change in N_e (hereafter referred to as the 'single-step-change analyses'). Using analytic likelihood calculations to fit such fully specified but minimally complex histories utilizes both allele frequency and linkage information and facilitates statistical comparison between species for the timing and magnitude of N_e change. We also applied a non-parametric method commonly used to visualize population size change, the pairwise sequentially Markovian coalescent (PSMC) (Li & Durbin, 2011), which is based on the distribution of pairwise differences in minimal samples of two haploid genomes. PSMC generates a more resolved picture of N_e change through time, including a qualitative assessment of the time at which populations have diverged. As the two methods make opposing assumptions, we will not attempt any formal comparison between them but rather compare inferences qualitatively. Both methods use different data properties and sampling strategies and, as a consequence, have contrasting limitations: while the single-step-change analyses—by design—cannot detect gradual N_e changes or resolve population histories involving multiple changes, PSMC is known to smooth out very sudden changes. PSMC is based on fewer samples which contain less information (i.e. fewer coalescence events) about N_e in the recent past (Li & Durbin, 2011). The two methods therefore complement each other and together provide a comprehensive picture of population history.

We address the following questions: (i) To what extent are the direction and timing of changes in N_e concordant across members of the oak gall parasitoid guild? Do species show evidence for simultaneous N_e change, suggesting concordant responses to a shared underlying driver, or are their demographic histories largely idiosyncratic as shown by Bunnefeld et al. (2018) for the timing of divergence and admixture between refugia? (ii) Do sudden changes in population size coincide with specific glacial or interglacial periods in the Pleistocene? (iii) In species that have an Eastern origin, do the most drastic demographic changes occur after the colonization of

Iberia, or are they shared with other refugial populations, suggesting that genetic diversity in Iberia largely reflects the size of the ancestral source populations?

2 | MATERIALS AND METHODS

2.1 | Samples and sequencing

We analysed whole-genome Illumina paired end resequencing data for six (haploid) male individuals in each of seven species of chalcid parasitoids (spanning five families): *Megastigmus dorsalis* and *M. stigmatizans* (Megastigmidae), *Torymus auratus* (Torymidae), *Ormyrus nitidulus* and *O. pomaceus* (Ormyridae), *Eurytoma brunniventris* (Eurytomidae) and *Cecidostiba fungosa* (Pteromalidae) (Table S1). The *M. dorsalis* individuals correspond to cryptic species 1 for this complex, as defined by Nicholls et al. (2010). For each species, we sampled five individuals from Iberia (the focal refugium) and one from Hungary (the Balkan refugium). Data for the Hungarian and two Iberian samples of each species were generated previously by Bunnefeld et al. (2018). We generated analogous data for the remaining three Iberian individuals using the protocols described by Bunnefeld et al. (2018). DNA was extracted from individual wasps using the Qiagen DNeasy kit. Individual Nextera genomic libraries were generated and sequenced on an Illumina HiSeq 2000 by the NERC Edinburgh Genomics facility, UK. Raw reads were deposited at the SRA (PRJEB20883). Mean coverage per haploid individual ranged from 3.5x to 18.5x. Raw reads were mapped back to reference genomes assembled by Bunnefeld et al. (2018) using BWA (0.7.15-r1140) (Li & Durbin, 2009), duplicates were marked with *picard* (V2.9.0) MarkDuplicates (broadinstitute.github.io/picard/), and variants were called using *Freebayes* (v1.1.0-3-g961e5f3) (<https://github.com/ekg/freebayes>) with a minimum base quality of 10 and a minimum mapping quality of 20 (see Bunnefeld et al. (2018) for further details).

2.2 | Inferring single-step-change models in population size

We fitted a model of a single, instantaneous step change in N_e using the framework for blockwise likelihood calculations developed by Lohse et al. (2011). The single-step-change model includes three parameters: the scaled mutation rate $\theta = 4N_0\mu l$ (where N_0 is the current N_e and l is block length); T , the time of N_e change measured in $2N_e$ generations; and $\lambda = N_0/N_1$, the relative magnitude of the N_e change (where N_1 denotes the N_e prior to the step change).

Following Bunnefeld et al. (2015), we summarized sequence variation in short blocks of a fixed length l by the (folded) blockwise site frequency spectrum (bSFS) using the script *block_cutter_vcf.py* (https://github.com/KLohse/BunnefeldEtAL_2018/block_cutter_vcf.py). Given our sampling scheme of $n = 5$ haploid males, the bSFS consists of counts of two types of variants: those for which the

minor allele occurs once or twice in the sample. The probability of observing a particular set of mutations in a block can be computed analytically as a higher-order derivative of the generating function of genealogies (Lohse et al., 2011). The product of probabilities of bSFS configurations across blocks can be interpreted as the composite likelihood (CL) of the model. We maximized $\ln CL$, i.e. *model support*, in *Mathematica* (Wolfram Research, 2016) using the function *NMaximize* (Data S1).

To generate blockwise data for each species, we applied the same quality filters used for calling SNPs to all sites; that is, we identified regions of the genome with a base quality >10 and mapping quality >20 in each individual from bam files by the CallableLoci walker of GATK (v3.4) (Van der Auwera et al., 2013). Only regions meeting these criteria in all five Iberian individuals were included in further analyses. Custom scripts were used to partition the data into blocks of a fixed length l of callable sites. l was chosen to be inversely proportional to the pairwise genetic diversity of each species (Table 2) such that blocks, each capturing a random configuration of closely linked variants had an average two pairwise differences between samples. This ensures that the information content per block, as well as any effect of intra-block recombination, is consistent across species. Blocks with a physical span (including non-callable sites) of >2l or more than five 'None' (uncalled) sites were removed.

We assessed support for a step change in N_e relative to the (nested) null model of constant N_e by generating parametric bootstrap replicates with the coalescent simulator *msprime* (Kelleher et al., 2016). For each species, 100 replicate data sets were simulated under the null model assuming estimates of recombination inferred by Bunnefeld et al. (2018) (Table S3). Each data set had the same total length as the real data (after filtering) and was partitioned into 5,000 windows of sequences (for computational efficiency). Both a null model of constant N_e and a step-change model were fitted to each replicate, and the 95% quantile of the difference in support between models was compared to that of the real data. Confidence intervals (CIs) of parameter estimates were obtained via an analogous parametric bootstrapping procedure: we simulated 100 data sets with recombination under the inferred step-change model and fitted that model to each simulation replicate. CIs were obtained as 2.5% and 97.5% quantiles of the distribution of parameter estimates

and were centred around the point estimates of parameters obtained from the real data.

2.3 | Reconstructing ancestral population size with PSMC

The pairwise sequentially Markovian coalescent (PSMC) (Li & Durbin, 2011) was used to infer a history of population size change for each species. PSMC is a non-parametric method that reconstructs a trajectory of past N_e from the density of pairwise differences along the genome via a hidden Markov model in which the hidden states are pairwise coalescence times, the distribution of which is used to estimate N_e in discrete time intervals. In each species, the two Iberian individuals with the greatest average read depth were used as the focal pair. Pileup files were generated using *samtools mpileup* (version 0.1.19) (Li et al., 2009), and a consensus sequence (fastq file) per individual was generated using the *bcftools* utility *vcf2fq*. Regions covered in both individuals were combined into fastq files using *seqtk mergefa* (<https://github.com/lh3/seqtk>) and converted into PSMC input files. PSMC, by default, discretizes pairwise alignments into blocks of 100 bp which are encoded as variant if they contain at least one variant. While this makes analyses of large genomes with low diversity (e.g. humans) computationally efficient, this discretization is too coarse when considering more diverse genomes where the chance of several pairwise differences occurring in the same 100 bp block is non-negligible. We investigated the effect of varying block length (100, 50, 25 and 1 bp) on N_e inference for the most diverse (*E. brunniventris*, $\pi = 0.0071$) and the least diverse (*M. stigmatizans*, $\pi = 0.00067$) species in our set. As expected, population trajectories showed higher N_e and were pushed back in time with smaller block lengths (Figure S1). We chose a block length of 25 bp for all analyses which minimizes these biases without excessive computational cost.

We inferred 30 free interval parameters across 64 time intervals (with the option -p '28*2+3+5'). The maximum coalescence time (-t) was set to 5, the initial value of θ/ρ (-r) to 1. 100 bootstraps were performed for each run. Times of peak N_e and values of N_e in each time interval were considered to be different between species if

TABLE 2 Summaries of PSMC trajectories for Iberian populations of seven species of chalcid parasitoid wasps. Times are given in thousands of years ago (kya). 95% CIs of peak N_e times are shown in brackets. The maximum N_e is scaled relative to π as $\text{Max}[N_e]/N_{e,\pi}$

Species	π	Peak $N_e \times 10^3$	Peak N_e time (kya)	Split time (kya)	Relative Max $[N_e]$
<i>Torymus auratus</i>	0.0064	835	78 (74–82)	n/a	1.81
<i>Megastigmus stigmatizans</i>	0.00067	65	47 (42–52)	37	1.34
<i>Megastigmus dorsalis</i>	0.0018	139	35 (31–39)	128	1.07
<i>Ormyrus nitidulus</i>	0.0016	157	114 (104–124)	132	1.36
<i>Ormyrus pomaceus</i>	0.0028	275	35 (30–40)	681	1.36
<i>Eurytoma brunniventris</i>	0.0071	831	46 (36–56)	n/a	1.62
<i>Cecidostiba fungosa</i>	0.0032	213	25 (12–38)	267	0.92

there was no overlap in bootstrap replicates. To be able to compare the magnitude of inferred N_e change between PSMC and single-step-change analyses, we normalized the maximum N_e inferred by each method by the long-term average N_e estimated from π Nei, 1972 (Table 2) as $\hat{N}_{e,\pi} = \pi/(4\mu)$ and computed the following measure of N_e change: $\text{Max}[N_e]/\hat{N}_{e,\pi}$. Unlike the size of the step change (λ), this measures the maximum N_e relative to the average over the entire history of a sample and is therefore expected to be smaller than λ .

2.4 | Calibrating the timing of events

Time estimates were converted into years assuming a mutation rate of 3.46×10^{-9} mutations per base per generation estimated for *Drosophila melanogaster* (Keightley et al., 2009). We assumed two generations per year for all species with the exception of *M. stigmatizans* which has a single generation per year (Stone et al., 2012). While this calibration allows comparisons with the estimates obtained by Bunnefeld et al. (2018) (calibrated in the same way), these absolute times are likely underestimates and should be treated with caution (given that we use a spontaneous mutation rate estimate and assume selective neutrality). We stress, however, that comparing the relative timing of demographic events across this set of parasitoids only relies on the assumption of the same per generation mutation rate across species rather than any absolute calibration.

2.5 | Cross-population PSMC analyses

To test whether population size changes in each species occurred before or after the divergence between the Iberian and Balkan refuge populations, we compared the PSMC trajectories of Iberian pairs and cross-population pairs (one Iberian and one Hungarian individual). Any divergence between Iberian and the Balkan refuge populations should be visible as a separation between within-population (Iberian pair) and cross-population (Iberia-Balkan pair) PSMC trajectories. Specifically, in the absence of significant post-divergence gene flow, we expect N_e estimates for the cross-population pairs to increase from the time of the population split due to the accumulation of genetic differences between the populations. Thus, population size changes that occurred in an ancestral population (potentially outside Iberia) should predate the divergence of within and cross-population trajectories. In contrast, we would expect demographic events unique to the Iberian population to happen after divergence of the within- and cross-population PSMC trajectories. We also compared our PSMC divergence time estimates to those made by Bunnefeld et al. (2018) under explicit models of divergence and admixture.

2.6 | Potential population structure in Iberia

Population structure within any assumed panmictic population can confound inferences of past demography (Bunnefeld et al.,

2015; Gattepaille et al., 2013). In particular, N_e estimates may be inflated due to divergence between demes (Gattepaille et al., 2013). Likewise, when samples are taken from the same deme, the (structured) coalescent generates a mixture of very recent within-deme ancestry and older ancestry resulting from migration between demes (Wakeley, 2008), which can mimic the signatures of a bottleneck (Bunnefeld et al., 2015). To test for population structure within Iberia, we repeated PSMC analyses on every pairwise combination of our five individuals. In a structured population, and given that the sampling locations were widely spaced across Iberia (Table S1), N_e trajectories are expected to differ between pairs of haploid males sampled from the same *versus* different sub-populations (analogous to the within- and between-population analyses involving Iberian and Hungarian samples).

2.7 | Sensitivity analyses

The assumption of no recombination within blocks (required to make the composite likelihood estimation of the single-step-change model tractable) potentially biases parameter estimates. Specifically, recombination may lead to a downward bias of N_e estimates (Bunnefeld et al., 2015; Wall, 2003). We used simulations in *msprime* (Kelleher et al., 2016) to quantify the effect of recombination on parameter estimates (Table S2). One million unlinked blocks of 586 bp (corresponding to the block length used for *O. nitidulus*) were simulated under a step-change model with different recombination rates and step sizes.

It is well known that both selective sweeps (Smith & Haigh, 1974) and background selection (Charlesworth et al., 1993) affect variation at neutral sites in the genome which, in turn, can bias demographic inference (Ewing & Jensen, 2016; Schrider et al., 2016). To explore the effect of selection on the single-step-change analyses, we fitted step-change models separately to blockwise data generated from genic and intergenic regions of the *O. nitidulus* genome. Genes were predicted *ab initio* with *Augustus* (Stanke & Morgenstern, 2005). *Nasonia vitripennis*, a chalcid parasitoid (family Pteromalidae), was used as a training set. If selection has a strong effect on demographic inference, estimates of θ are expected to be lower for genic compared to intergenic regions, as most selection tends to reduce diversity both at selective targets and linked regions of the genome (Smith & Haigh, 1974). Similarly, we would expect estimates for the time of the step change, T , to be biased towards the present in genic compared to intergenic regions.

3 | RESULTS

After filtering, the blockwise data sets ranged in total length from 54 to 151 Mb (Table S3). The pairwise alignments used as input for PSMC spanned a total length of 161 to 379 Mb (Table S3). Despite this difference in overall length, which is mainly due to missing data and the difference in sample size (two *versus* five individuals),

estimates of average pairwise diversity, as measured by π (Nei, 1972), agreed well between the two data sets (Table 1). Across species, π estimates spanned an order of magnitude, from 0.00067 in *M. stigmatizans* to 0.0071 in *E. brunneiventris*.

3.1 | Large changes in N_e detected in four species

Taking the results of the single-step-change and PSMC analyses across all seven parasitoid species together, four species (*M. stigmatizans*, *M. dorsalis*, *O. nitidulus* and *T. auratus*) show evidence for large (at least a factor of three) changes in population size during the Quaternary period (Figure 1). In these species, an instantaneous step change in N_e fits the blockwise data significantly better than a null model of a constant N_e . We infer a decrease in N_e towards the present ($\lambda < 1$) in three species (*M. stigmatizans*, *M. dorsalis* and *O.*

nitidulus) and an increase in one species (*T. auratus*) ($\lambda > 1$) (Figure 1 and Table 1). In all four species, the change in N_e in the PSMC trajectory (Figure 1) agrees both in direction and in coarse timescale with the inference under the single-step-change model. However, the N_e changes visible in the PSMC trajectory for these species are not equally abrupt. For example, *T. auratus* shows a gradual increase of N_e over a period of more than 200 ky, while the decreases in the PSMC trajectories of *M. dorsalis* and *M. stigmatizans* are comparatively sudden. PSMC trajectories and single-step-change analyses are also broadly similar for *O. pomaceus* and *C. fungosa*: a step change in N_e is supported in neither species, and PSMC suggests comparatively small population size changes for both (Table 1 and Table 2).

However, while we have resisted the temptation to attempt any formal comparison between PSMC trajectories and single-step-change analyses which would necessarily be *post hoc* and potentially misleading (given the opposing assumptions of these methods, see

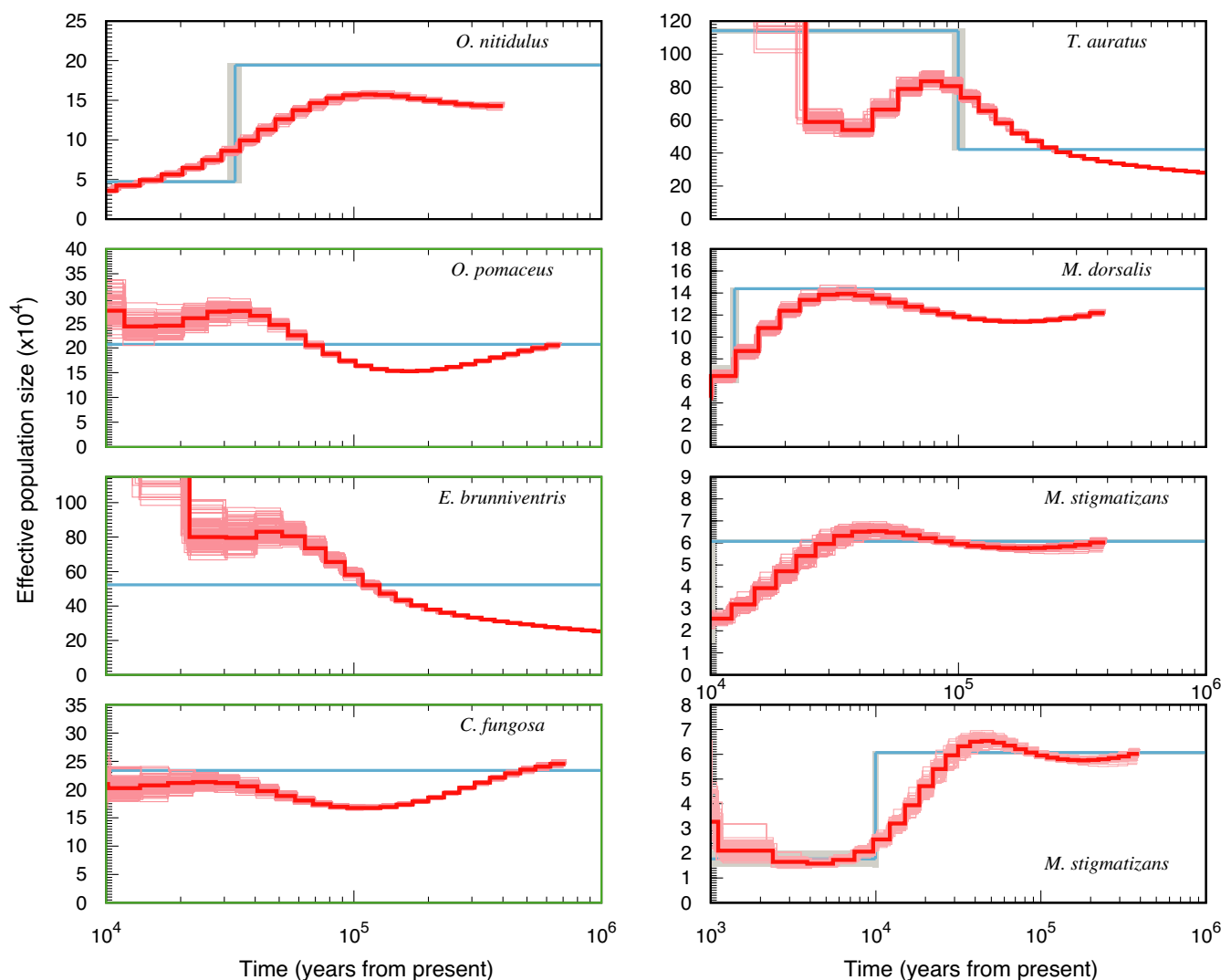


FIGURE 1 PSMC and maximum-composite likelihood estimates (MCLE) under the single-step-change model for Iberian populations of seven species of chalcid parasitoid wasps. PSMC estimates and bootstrap replicates are shown in dark red and pale red, respectively. Population sizes estimated using the composite likelihood step-change model are shown in blue with 95% CIs in grey. The three species with green plot frames are those for which a step-change model is not supported. Results for *M. stigmatizans* are also plotted on an alternative timescale to reveal recent N_e change (bottom right)

Discussion), inferences under the two methods disagrees qualitatively in at least two obvious ways:

First, with the exception of *M. stigmatizans*, the magnitude of N_e change inferred under the step-change model is greater than the relative magnitude of peak N_e in the PSMC trajectories (Figure 1, Table 1 and Table 2). Second, in one species, *E. brunniventris*, single-step-change and PSMC analyses are hard to reconcile: we found no significant support for a step change in this species, that is, we cannot reject a null model of a single fixed N_e . Yet, the PSMC trajectory indicates a substantial (but gradual) increase in N_e (Table 2). Additional analyses (see Discussion) suggest that demographic inferences for *E. brunniventris* may be affected by genetic structure and/or the low contiguity of its reference genome.

3.2 | No support for temporal concordance of population size change

The four species with support for a step change in N_e (*M. stigmatizans*, *M. dorsalis*, *O. nitidulus* and *T. auratus*) show no overlap in the estimated times of N_e change (Table 1). Using an insect spontaneous mutation rate (Keightley et al., 2009) to calibrate T estimates (see Methods for caveats), Iberian populations of *M. stigmatizans* and *M. dorsalis* most likely decreased in size at the start of the current interglacial (10 and 12.5 kya, respectively). In contrast, the decrease in N_e inferred for *O. nitidulus* most likely dates to the last glacial period (33 kya) and the increase in N_e in *T. auratus* at 100 kya falls to just after the end of the (Eemian) interglacial (Table 1). Formalizing this comparison, we conclude that a maximally complex model that assumes a unique time of N_e change for each of the four species fits significantly better than any scenario involving concordant/clustered times of N_e change.

Comparing times of N_e change inferred via PSMC across species is inherently problematic (see Discussion). However, when comparing the time of maximum N_e by computing the overlap in peak N_e intervals across bootstrap replicates, we find that the four taxa *M. stigmatizans*, *M. dorsalis*, *O. nitidulus* and *T. auratus* which show support for a single-step change at unique times also have non-overlapping peak N_e times (Table 1). Interestingly, the three species with no support for a single-step change (*O. pomaceus*, *C. fungosa* and *E. brunniventris*) and *M. dorsalis* show highly similar (overlapping CIs) times of peak N_e (Figure 2 and Table 2).

3.3 | Changes in N_e occur after the divergence of Iberian populations

In five species, the PSMC trajectories of within-population (Iberia) pairs diverge clearly from the cross-population (Iberia versus Hungarian) trajectories (Figure 3). In contrast, little divergence between within and cross-population PSMC trajectories is visible for *E. brunniventris* and none for *T. auratus* (Figure 3). In all three species for which the single-step-change analyses support a decline in

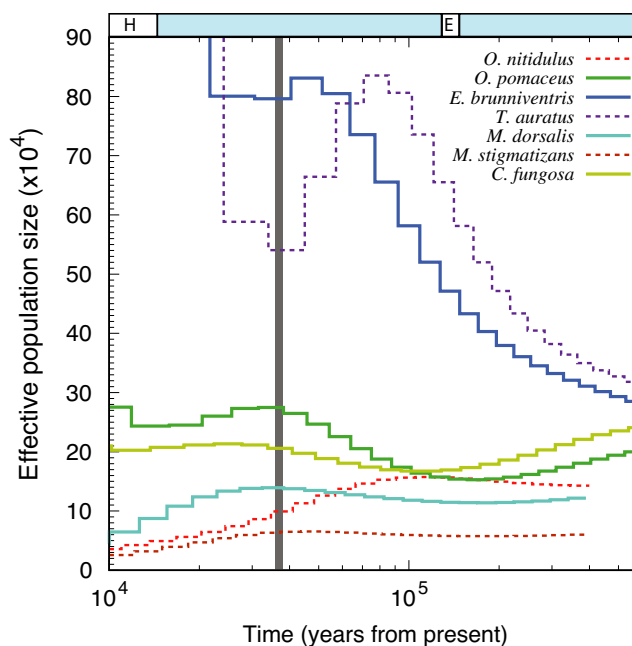


FIGURE 2 N_e trajectories inferred by PSMC for Iberian populations of seven species of parasitoids. The vertical grey bar indicates the overlap in the timing of peak N_e across 95% CIs computed from bootstrap replicates across four species (solid lines) (*M. dorsalis*, *O. pomaceus*, *C. fungosa* and *E. brunniventris*). The remaining three species (dashed lines) have unique peak N_e times. The time bar along the top of the figure shows (from left to right) the Holocene (H, 11.7 kya to present), the last glacial, the Eemian interglacial (E, 115–130 kya) and the penultimate interglacial

population size, the inferred time of N_e change is more recent than the divergence of within- and cross-population PSMC trajectories (Figure 3 and Table 2) and so must have occurred after Iberian and Balkan populations split. For *O. pomaceus*, *C. fungosa* and *M. stigmatizans*, the split times between Iberian and the Balkan populations inferred here *post hoc* by comparing PSMC trajectories are broadly compatible with the divergence estimates obtained by Bunnefeld et al. (2018) under explicit models of population divergence (Figure 3). In contrast, for both *M. dorsalis* and *O. nitidulus*, the divergence of within and between refuge PSMC trajectories substantially predates the split times inferred by Bunnefeld et al. (2018). In both cases, cross-population PSMC trajectories decrease after divergence between the Iberian and Hungarian populations, suggesting that these populations have been connected by a period of gene flow immediately following divergence which may not be detectable given the admixture pulse scenarios fitted by Bunnefeld et al. (2018).

3.4 | No evidence for structure within Iberia

We find little variation in PSMC trajectories between different Iberian pairs in almost all species (Figure S2), suggesting that our demographic inferences are unlikely to be influenced by population structure within Iberia. For all species except *E. brunniventris*, the

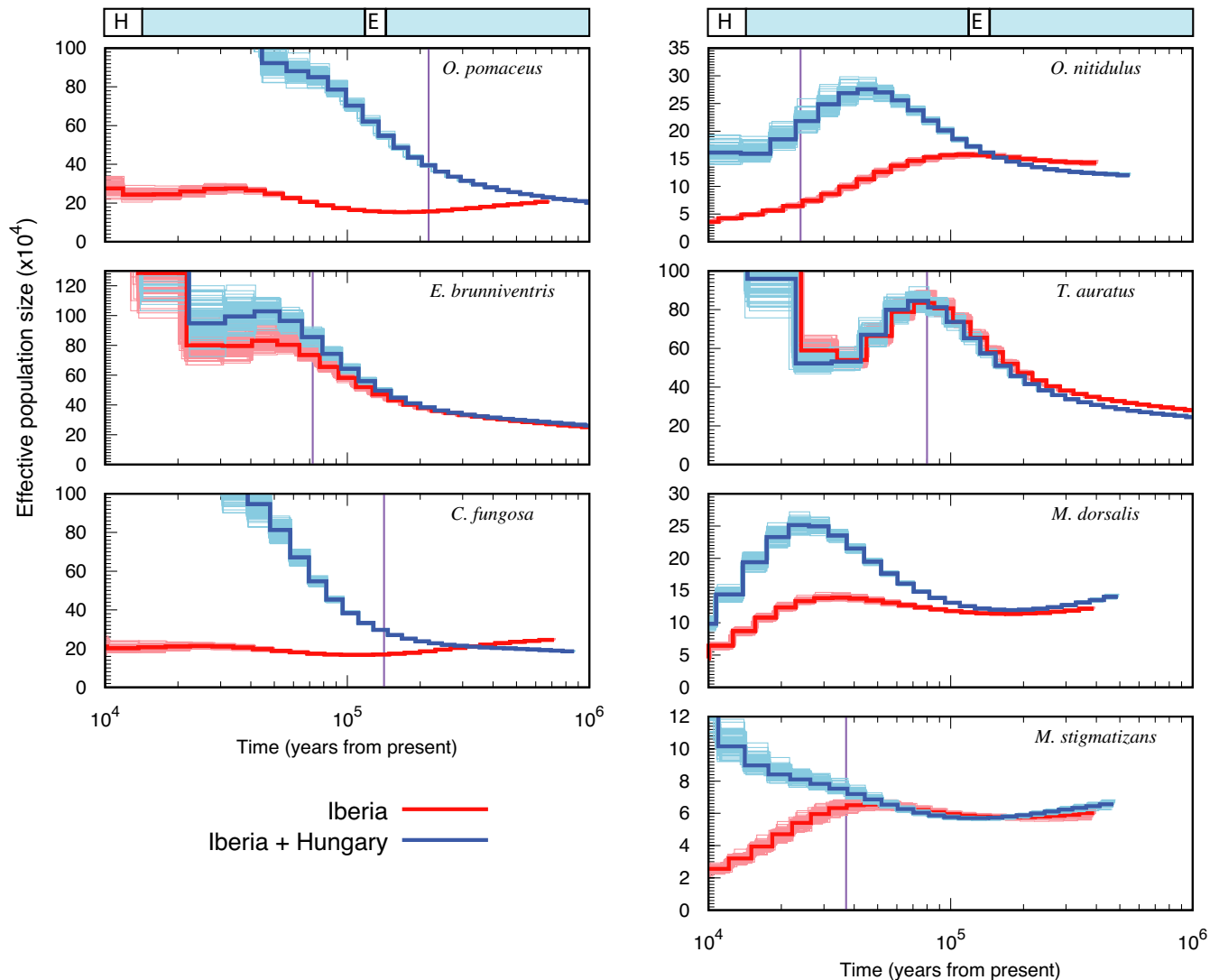


FIGURE 3 PSMC N_e trajectories for Iberian (red) and cross-population (Iberian vs Balkans) pairs of seven species of parasitoids. Population splits are visible as a divergence between the trajectories of within-population and cross-population pairs. Vertical bars indicate the split times estimated by Bunnefeld2018 using a MCL method based on the blockwise data. The split time estimated by Bunnefeld2018 for *M. dorsalis* is too recent (8 kya) to be visible in this figure. The time bar along the top of the figure shows (from left to right) the Holocene (H, 11.7 kya to present), the last glacial, the Eemian interglacial (E, 115–130 kya) and the penultimate interglacial

differences in PSMC trajectories between different Iberian sample pairs are similar to the differences across bootstrap replicates for the focal pair (Figure 1 and Figure S2) and so likely reflect coverage variation between individuals. *E. brunniventris* is the only species that showed clear variation in PSMC trajectories between Iberian pairs. While our additional analyses for *E. brunniventris* suggest other potential reasons for this finding (see Discussion), we cannot rule out that this variation is in part driven by population structure.

3.5 | Sensitivity analyses

Both demographic inference methods used here assume selective neutrality but make different simplifying assumptions about recombination: while PSMC approximates recombination as a Markov

process along the genome, the single-step-change analyses based on the bSFS assumes no recombination within blocks. To check the extent to which recombination and selection may bias parameter estimates, we fitted histories of a step change to data simulated with recombination. Our exploration of simulated data shows that both N_e and λ are underestimated with increasing recombination rate whereas the time of step change (T) is biased downwards when N_e declines towards the present ($\lambda < 1$) and upwards when N_e increases ($\lambda > 1$) (Table S2). These biases are an expected consequence of the shuffling of genealogical histories within blocks in the presence of recombination, which reduces the variance in bSFS configurations. Given a recombination rate of $r = 3 \times 10^{-9}$ (Table S3), our simulation check suggests that while T , the time of the step change may be underestimated by up to a factor of two, the ability to accurately estimate λ , the magnitude of the step change, is little affected.

To investigate the potential effect of selective constraint on the single-step-change analyses, we inferred single-step-change histories separately for genic and intergenic regions. Parameter estimates based on intergenic data for *O. nitidulus* were broadly similar to those obtained from the full data set (Table S2): while estimates of θ and T were slightly lower and higher, respectively, for the full data set, as would be expected as a result of selective constraint, the estimate of λ was little affected.

4 | DISCUSSION

We analysed genome-wide sequence variation using two contrasting inference approaches to test whether and how demographic histories vary within a guild of insect parasitoids in a single Pleistocene refugium. We find evidence for drastic declines in population size in three out of seven species (*M. stigmatizans*, *M. dorsalis*, *O. nitidulus*) and a large increase in population size in one (*T. auratus*). Interestingly, these four species encompass a mixture of longitudinal expansion histories: while Bunnefeld et al. (2018) inferred that *M. dorsalis* and *O. nitidulus* expanded into Iberia from the East, *M. stigmatizans* most likely expanded out of Iberia and *T. auratus* showed no longitudinal expansion signal. This suggests that the history of population size change in Iberia is not determined in any obvious or simple way by whether Iberia was the end point of an initial expansion into Europe or not. Contrary to any simple scenario of guild-wide temporal concordance in responses to Pleistocene climatic events, we find that population size changes of species in this guild differ markedly both in direction and in timescale. In fact, our single-step-change analyses reveal significant support for maximal temporal discordance; that is, each of the four species in this parasitoid guild that show evidence for a sudden change in N_e has a unique time (Table 1).

Thus, our main result of temporally discordant N_e change within Iberia mirrors the finding of temporal discordance by Bunnefeld et al. (2018) in the timing of divergence and admixture for this guild on a continental scale. One could argue that the signal of temporal discordance may simply be an artefact of applying an oversimplified step-change model, which is unlikely to capture the subtleties of real population size change. However, the fact that we also find evidence for temporal discordance when visualizing N_e change using PSMC, which makes minimal simplifying assumptions about the shape of past demographic change, suggests a genuine lack of signal for temporal concordance in this parasitoid guild across a range of spatio-temporal scales. This general finding mirrors results of other comparative studies on sets of co-distributed taxa in Europe (Ebdon et al., 2021) and the Americas (Burbrink et al., 2016; Dasmahapatra et al., 2010).

It is interesting that the only potential signal of temporal concordance we find is for four species that show the smallest change in past population sizes and which overlap in the time of peak N_e (Figure 2). While this apparent congruence may be due to a shared background demography, the signatures of which are masked in species with more drastic changes in N_e , we cannot rule out alternative

explanations. In particular, the fact that PSMC assumes selective neutrality is problematic given that insect genomes are more compact than mammalian genomes (Li & Graur, 1991), and so more susceptible to the effects of linked selection. Schrider et al. (2016) have shown that selective sweeps can generate troughs in PSMC trajectories while background selection has been shown to lead to erroneous inference of population growth (Ewing & Jensen, 2016). We therefore cannot rule out the possibility that congruent peaks in N_e are an artefact of selective effects which, assuming similar genome composition, mutation and recombination rates, may lead to similarly distorted PSMC inferences. However, the fact that we have reconstructed strikingly different PSMC trajectories, most of which differ markedly from the selection-induced PSMC trajectories of Schrider et al. (2016) (in that they show large declines rather than increases in N_e towards the present), suggests that it is unlikely that linked selection is the main driver of the inferred N_e changes. For the single-step-change analyses, we find little difference in parameter estimates when analysing all data or just intergenic regions (Table S2), which again argues against a major effect of selection on our inferences. Furthermore, one would expect genomes with a shorter map length (physical length \times recombination rate, see Table S3) to be disproportionately affected by linked selection (Mackintosh et al., 2019; Smith & Haigh, 1974). However, if anything, we observe the opposite pattern: the two species with shortest map lengths (*O. nitidulus* and *C. fungosa*) show less pronounced N_e change than the two species with the longest map length (*M. stigmatizans* and *M. dorsalis*).

4.1 | Reconciling single-step-change and PSMC analyses

PSMC and single-step-change analyses exploit different aspects of the data and differ drastically in sampling schemes (contiguous pairwise alignments vs short blocks sampled across five individuals) and underlying assumptions: while the blockwise composite likelihood framework fits a single instantaneous step change in N_e and infers the timing of this event, PSMC does not estimate a time parameter *per se*, but rather imposes an arbitrary discretization of time and reconstructs population size change as a continuous trajectory. It is reassuring that despite these fundamental differences, both methods yield broadly congruent conclusions: the four species for which the single-step-change analyses diagnose an abrupt change in N_e also show PSMC trajectories with large N_e changes in the same direction and at similar times. The greater magnitude of N_e change for the single-step-change compared to the PSMC analyses may be a consequence of the fact that larger samples used to fit the single-step-change model contain more information about recent demography than a pair of lineages.

In general, one may view the fact that PSMC is essentially assumption-free as an advantage over model-based inference, because it enables a straightforward visualization of past N_e change. Similarly, comparing PSMC trajectories between pairs of individuals and populations allows qualitative assessment of likely periods of

divergence and admixture (Figure 3). However, the flip-side of this flexibility is that PSMC provides no obvious route for quantitatively testing (necessarily) simple demographic hypotheses across species. Furthermore, cross-species comparisons of PSMC trajectories are problematic even when they are focused on a single clearly defined summary such as the interval of peak N_e (as we have done here), because the time discretization PSMC imposes depends on the rate of coalescence, and so differs between species.

4.2 | Changes in population size coincide with late Pleistocene climatic transitions

While we emphasize that absolute time estimates need to be interpreted with caution, our estimates of the timing of drastic N_e changes coincide broadly with climatic events in the Quaternary: the start of the Holocene around 11 kya and the end of the Eemian interglacial around 106 kya (Figure 3). Previous studies on the parasitoid oak gall wasp community based on the same calibration have inferred an increase in gene flow between refugia during the same time periods (Bunnefeld et al., 2018; Lohse et al., 2010, 2012). For example, the cluster of peak N_e times coincides with a large community-wide peak in the frequency of admixture between refugia inferred by Bunnefeld et al. (2018). Similarly, the large increase in N_e inferred for *T. auratus* at the end of the Eemian interglacial period coincides with the divergence of refugial populations inferred for this species previously (Bunnefeld et al., 2018; Stone et al., 2012). It seems plausible that both events are associated with the geographic expansion of suitable oak habitat across refugial barriers during these times (Brewer et al., 2002; Petit et al., 2002), which may have triggered range expansions in both host gall wasps and their associated parasitoids.

4.3 | Population structure within Iberia and gene flow from Eastern refugia

Population structure within southern European refugia has previously been demonstrated for some species, and it has been suggested that given its complex topography Iberia should be considered a mosaic of multiple micro-refugia rather than a single entity (Feliner, 2011; Hearn et al., 2014). However, our PSMC results suggest a complete lack of population structure in six out of seven species in the parasitoid guild (Figure S2) and imply high gene flow within Iberia. This is perhaps unsurprising given that gall wasp-associated parasitoid wasps (and other chalcids) are able to disperse long distances even across patchy habitats and host distributions (Compton et al., 2000; Hayward & Stone, 2006).

Our estimates of population split times based on comparisons of within- and cross-population PSMC trajectories agree broadly with those of Bunnefeld et al. (2018) for most species. *M. dorsalis* and *O. nitidulus*, the two species that show the least agreement with past estimates, both show decreases in cross-population N_e after

divergence that are compatible with ongoing gene flow between refugia. The model space considered by Bunnefeld et al. (2018) was limited to histories involving a single discrete burst of instantaneous admixture between refugial populations, with limited potential to detect periods of continuous post-divergence gene flow.

4.4 | *Eurytoma brunniventris* is an outlier

Eurytoma brunniventris is an outlier in our results in several ways: it is the species with the highest genetic diversity, shows signals of population structure within Iberia and is the only species for which our two inference approaches disagree. While the single-step-change analyses give no support for a change in N_e , the PSMC trajectory shows a steady increase. To explore the sensitivity of the single-step-change analysis to detect gradual changes in population size, we simulated 100 replicate data sets (assuming the same size and block length as the real data) under the gradual change in N_e inferred via PSMC for *E. brunniventris*. We find that in each case, a single-step-change history fits the data significantly better (using a parametric bootstrap analogous to that performed on the real data, see Methods) than a null model of constant N_e , indicating that the single-step-change analysis is indeed sensitive to gradual increases in population size. A possible explanation for the discrepancy between the PSMC and single-step-change analyses for *E. brunniventris* is that its genome assembly, the least contiguous among our set of taxa (Table S3), is too fragmented for reliable PSMC inference. Interestingly, both a history of constant N_e and a single-step-change model give a poor fit to the observed frequency of bSFS configurations in *E. brunniventris*. In particular, *E. brunniventris* shows an excess of both monomorphic blocks and blocks with a large number of variants (Figure S3), suggesting that its history is not well approximated by any model that assumes a single panmictic population. The lack of divergence of within- and between-population PSMC trajectories for *E. brunniventris* would be compatible with substantial gene flow between the Iberian and Hungarian populations (Figure 3). Alternatively, *E. brunniventris*—an extreme generalist attacking a wide range of oak gall wasp hosts (Askew et al., 2013)—may harbour genetic structure as a result of recent divergence into cryptic host races (Nicholls et al., 2018). However, in the absence of a better reference genome and larger sample sizes, it remains unclear to what extent the disagreement between PSMC and single-step-change analyses for *E. brunniventris* is indicative of a more complex history.

4.5 | Outlook

A general question for geographically widespread communities is the extent to which component species continue to interact during the assembly process (Agosta & Klemens, 2008; Janzen, 1985; Ricklefs, 2015). Where communities are characterized by strong dependencies between species, such as specific trophic or symbiotic interactions, we might expect co-dispersal and coupled population

dynamics to result in similar demographic histories (e.g. Gaume et al., 2000); this scenario is compatible with ongoing selective effects of species on each other and potential coevolution (Hall et al., 2020; Wade, 2007).

In contrast, we might expect much lower demographic concordance for members of communities characterized by weaker and less specific species interactions and only diffuse coevolution (Hall et al., 2020). This is the pattern emerging for the parasitoid assemblages attacking oak gall wasps. Though comprising a consistent set of interacting taxa spanning thousands of kilometres of longitude, the component species show highly diverse histories of range expansion (Bunnefeld et al., 2018) and changes in population size. This finding is consistent with the ability of most of the parasitoid species to attack multiple gall wasp hosts (Askew et al., 2013), weakening both direct interactions between the trophic levels and competitive interactions between parasitoid species. While there is strong evidence that host gall wasp traits structure-associated assemblages of parasitoid enemies (Bailey et al., 2009) and of parasitoid specializations for exploitation of particular hosts (Weinersmith et al., 2017), there is little evidence for specific coevolution. The population histories of very few other multitrophic communities have been explored, and the extent to which the demographic diversity within the gall wasp system is typical of parasitoid–host and other biological systems remains unknown. Work on the arthropod communities associated with the pitcher plant *Sarracenia alata* (Satler & Carstens, 2017) also shows variation in component species histories; further, this study suggests that more ecologically dependent associates show closer demographic and phylogeographic concordance with the host plant than less ecologically dependent species. An obvious question is whether any of the variation in patterns of N_e change in our sampled parasitoid species can be attributed to variation in ecological traits (Papadopolou & Knowles, 2016). Intriguingly, the four parasitoid species with support for a step change (in either direction) have a narrower host range (Askew et al., 2013) and a lower ancestral N_e than those for which a null model of constant N_e could not be rejected (Table S1). Although the number of species studied here is clearly insufficient for any statistical test of an association between host range and demographic history, these trends are compatible with the idea that ecologically specialist species (whose biology is critically dependent on interactions with a small number of other taxa) experience greater and/or more frequent changes in N_e than generalists (whose demography is less tightly coupled to abundance of any specific interaction) (Östergård & Ehrlén, 2005; Rand & Tscharnke, 2007). Larger samples of species, incorporating wide diversity in host number and other relevant traits (such as dispersal ability) are required to assess the extent to which life history traits and demographic histories are correlated. Given that the genomes of parasitoid wasp can be sampled in a haploid state (by targeting males), it will be fascinating to test how much more signal about the demographic past of ecological communities can be extracted using a new generation of inference approaches that reconstruct the sequence of correlated genealogies directly from the data (Kelleher et al., 2019; Speidel et al., 2019).

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AUTHOR CONTRIBUTIONS

KL and GS designed the project; WW analysed the sequence data with contributions from KL; all authors wrote the manuscript.

DATA AVAILABILITY STATEMENT

- Raw reads have been deposited in the European Nucleotide Archive (ENA) (ERP023079) and the SRA (PRJEB20883)
- Genome assemblies are deposited in the ENA (PRJEB27189 and ERP109243)
- *Mathematica* notebook and blockwise data are available as Supporting Information

ORCID

Graham N. Stone  <https://orcid.org/0000-0002-2737-696X>

Konrad Lohse  <https://orcid.org/0000-0001-9918-058X>

REFERENCES

- Agosta, S. J., & Klemens, J. A. (2008). Ecological fitting by phenotypically flexible genotypes: implications for species associations, community assembly and evolution. *Ecology Letters*, 11(11), 1123–1134.
- Askew, R. R., Melika, G., Pujade-Villar, J., Schönrogge, K., Stone, G. N., & Nieves-Aldrey, J. L. (2013). Catalogue of parasitoids and inquilines in cynipid oak galls in the West Palaearctic. *Zootaxa*, 3643(1), 1–133.
- Avise, J. C., Arnold, J., Ball, R. M., Bermingham, E., Lamb, T., Neigel, J. E., Reeb, C. A., & Saunders, N. C. (1987). Intraspecific phylogeography: The mitochondrial DNA bridge between population genetics and systematics. *Annual Review of Ecology and Systematics*, 18(1), 489–522.
- Bailey, R., Schönrogge, K., Cook, J. M., Melika, G., Csóka, G., Thuróczy, C., & Stone, G. N. (2009). Host niches and defensive extended phenotypes structure parasitoid wasp communities. *PLOS Biology*, 7(8), 1–12.
- Beichman, A. C., Huerta-Sanchez, E., & Lohmueller, K. E. (2018). Using genomic data to infer historic population dynamics of nonmodel organisms. *Annual Review of Ecology, Evolution, and Systematics*, 49(1), 433–456.
- Bermingham, E., & Avise, J. C. (1986). Molecular zoogeography of freshwater fishes in the southeastern United States. *Genetics*, 113(4), 939–965.
- Brewer, S., Cheddadi, R., de Beaulieu, J., & Reille, M. (2002). The spread of deciduous *Quercus* throughout Europe since the last glacial period. *Forest Ecology and Management*, 156(1–3), 27–48.
- Bunnefeld, L., Frantz, L. A. F., & Lohse, K. (2015). Inferring bottlenecks from genome-wide samples of short sequence blocks. *Genetics*, 201(3), 1157–1169.
- Bunnefeld, L., Hearn, J., Stone, G. N., & Lohse, K. (2018). Whole-genome data reveal the complex history of a diverse ecological community. *Proceedings of the National Academy of Sciences of the United States of America*, 115(28), E6507–E6515.

- Burbrink, F. T., Chan, Y. L., Myers, E. A., Ruane, S., Smith, B. T., & Hickerson, M. J. (2016). Asynchronous demographic responses to pleistocene climate change in eastern nearctic vertebrates. *Ecology Letters*, 19(12), 1457–1467.
- Charlesworth, B., Morgan, M. T., & Charlesworth, D. (1993). The effect of deleterious mutations on neutral molecular variation. *Genetics*, 134(4), 1289–1303.
- Comps, B., Gömöry, D., Letouzey, J., Thiébaud, B., & Petit, R. J. (2001). Diverging trends between heterozygosity and allelic richness during postglacial colonization in the European beech. *Genetics*, 157(1), 389–397.
- Compton, S. G., Ellwood, M. D. F., Davis, A. J., & Welch, K. (2000). The Flight Heights of Chalcid Wasps (Hymenoptera, Chalcidoidea) in a Lowland Bornean Rain Forest: Fig Wasps are the High Fliers. *Biotropica*, 32(3), 515–522.
- Dasmahapatra, K. K., Lamas, G., Simpson, F., & Mallet, J. (2010). The anatomy of a 'suture zone' in amazonian butterflies: a coalescent-based test for vicariant geographic divergence and speciation. *Molecular Ecology*, 19(19), 4283–4301.
- Ebdon, S., Laetsch, D. R., Dapporto, L., Hayward, A., Ritchie, M. G., Dincă, V., Vila, R., & Lohse, K. (2021). The pleistocene species pump past its prime: evidence from European butterfly sister species. *Molecular Ecology*, 30, 3575–3589. <https://doi.org/10.1111/mec.15981>
- Ewing, G. B., & Jensen, J. D. (2016). The consequences of not accounting for background selection in demographic inference. *Molecular Ecology*, 25(1), 135–141.
- Feliner, G. N. (2011). Southern European glacial refugia: A tale of tales. *Taxon*, 60(2), 365–372.
- Gattepaille, L. M., Jakobsson, M., & Blum, M. G. B. (2013). Inferring population size changes with sequence and SNP data: lessons from human bottlenecks. *Heredity*, 110(5), 409–419.
- Gaume, L., Matile-Ferrero, D., & McKey, D. (2000). Colony foundation and acquisition of coccid trophobionts by aphomomyrmex afer (formicinae): co-dispersal of queens and phoretic mealybugs in an ant-plant-homopteran mutualism? *Insectes Sociaux*, 47(1), 84–91. <https://doi.org/10.1007/s000400050013>.
- Gutenkunst, R. N., Hernandez, R. D., Williamson, S. H., Bustamante, C. D., & Stephan, W. (2009). Inferring the joint demographic history of multiple populations from multidimensional SNP frequency data. *PLoS Genetics*, 5(10), e1000695.
- Hall, A. R., Ashby, B., Bascompte, J., & King, K. C. (2020). Measuring coevolutionary dynamics in species-rich communities. *Trends in Ecology & Evolution*, 35(6), 539–550.
- Hayward, A., & Stone, G. N. (2006). Comparative phylogeography across two trophic levels: the oak gall wasp *Andricus kollari* and its chalcid parasitoid *Megastigmus stigmatizans*. *Molecular Ecology*, 15(2), 479–489.
- Hearn, J., Stone, G. N., Bunnefeld, L., Nicholls, J. A., Barton, N. H., & Lohse, K. (2014). Likelihood-based inference of population history from low-coverage *de novo* genome assemblies. *Molecular Ecology*, 23(1), 198–211.
- Hewitt, G. M. (1996). Some genetic consequences of ice ages, and their role in divergence and speciation. *Biological Journal of the Linnean Society*, 58(3), 247–276.
- Hewitt, G. (2000). The genetic legacy of the Quaternary ice ages. *Nature*, 405(6789), 907–913.
- Hickerson, M., Carstens, B., Cavender-Bares, J., Crandall, K., Graham, C., Johnson, J., Rissler, L., Victoriano, P., & Yoder, A. (2010). Phylogeography's past, present, and future: 10 years after avise, 2000. *Molecular Phylogenetics and Evolution*, 54(1), 291–301.
- Hofreiter, M., & Stewart, J. (2009). Ecological change, range fluctuations and population dynamics during the pleistocene. *Current Biology*, 19(14), R584–R594.
- Janzen, D. (1985). On ecological fitting. *Oikos*, 45(3), 308–310.
- Keightley, P. D., Trivedi, U., Thomson, M., Oliver, F., Kumar, S., & Blaxter, M. L. (2009). Analysis of the genome sequences of three *Drosophila melanogaster* spontaneous mutation accumulation lines. *Genome Research*, 19(7), 1195–1201.
- Kelleher, J., Etheridge, A. M., & McVean, G. (2016). Efficient coalescent simulation and genealogical analysis for large sample sizes. *PLoS Computational Biology*, 12(5), 1–22.
- Kelleher, J., Wong, Y., Wohns, A. W., Fadil, C., Albers, P. K., & McVean, G. (2019). Inferring whole genome histories in large population dataset. *Nature Genetics*, 51(9), 1330–1338.
- Leaché, A. D., Crews, S. C., & Hickerson, M. J. (2007). Two waves of diversification in mammals and reptiles of baja california revealed by hierarchical bayesian analysis. *Biology Letters*, 3(6), 646–650.
- Li, H., & Durbin, R. (2009). Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics*, 25(14), 1754–1760.
- Li, H., & Durbin, R. (2011). Inference of human population history from individual whole-genome sequences. *Nature*, 475(7357), 493–496.
- Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., Marth, G., Abecasis, G., & Durbin, R. (2009). The Sequence Alignment/Map format and SAMtools. *Bioinformatics*, 25(16), 2078–2079.
- Li, W. H., & Graur, D. (1991). *Fundamentals of molecular evolution*. Sinauer Associates.
- Lohse, K., Barton, N., Melika, G., & Stone, G. (2012). A likelihood-based comparison of population histories in a parasitoid guild. *Molecular Ecology*, 21, 4605–4617.
- Lohse, K., Harrison, R. J., & Barton, N. H. (2011). A general method for calculating likelihoods under the coalescent process. *Genetics*, 189(3), 977–987.
- Lohse, K., Sharanowski, B., & Stone, G. N. (2010). Quantifying the pleistocene history of the oak gall parasitoid *Cecidostiba fungosa* using twenty intron loci. *Evolution*, 64(9), 2664–2681.
- Mackintosh, A., Laetsch, D. R., Hayward, A., Charlesworth, B., Waterfall, M., Vila, R., & Lohse, K. (2019). The determinants of genetic diversity in butterflies. *Nature Genetics*, 10(1), 3466.
- Nei, M. (1972). Genetic distance between populations. *The American Naturalist*, 106(949), 283–292.
- Nicholls, J. A., Challis, R. J., Mutun, S., & Stone, G. N. (2012). Mitochondrial barcodes are diagnostic of shared refugia but not species in hybridizing oak gallwasps. *Molecular Ecology*, 21(16), 4051–4062.
- Nicholls, J. A., Preuss, S., Hayward, A., Melika, G., Csöka, G., Nieves-Aldrey, J.-L., Askew, R. R., Tavakoli, M., Schönrogge, K., & Stone, G. N. (2010). Concordant phylogeography and cryptic speciation in two Western Palaearctic oak gall parasitoid species complexes. *Molecular Ecology*, 19(3), 592–609.
- Nicholls, J. A., Schönrogge, K., Preuss, S., & Stone, G. N. (2018). Partitioning of herbivore hosts across time and food plants promotes diversification in the megastigmus dorsalis oak gall parasitoid complex. *Ecology and Evolution*, 8(2), 1300–1315.
- Östergård, H., & Ehrlén, J. (2005). Among population variation in specialist and generalist seed predation the importance of host plant distribution, alternative hosts and environmental variation. *Oikos*, 111(1), 39–46.
- Papadopoulou, A., & Knowles, L. L. (2016). Toward a paradigm shift in comparative phylogeography driven by trait-based hypotheses. *Proceedings of the National Academy of Sciences USA*, 113(29), 8018–8024.
- Petit, R. J., Brewer, S., Bordács, S., Burg, K., Cheddadi, R., Coart, E., Cottrell, J., Csaikl, U. M., van Dam, B., Deans, J. D., Espinel, S., Fineschi, S., Finkeldey, R., Glaz, I., Goicoechea, P. G., Jensen, J. S., König, A. O., Lowe, A. J., Madsen, S. F., ... Kremer, A. (2002). Identification of refugia and post-glacial colonisation routes of European white oaks based on chloroplast DNA and fossil pollen evidence. *Forest Ecology and Management*, 156(1–3), 49–74.
- Rand, T. A., & Tscharnkte, T. (2007). Contrasting effects of natural habitat loss on generalist and specialist aphid natural enemies. *Oikos*, 116(8), 1353–1362.
- Ricklefs, R. E. (2015). Intrinsic dynamics of the regional community. *Ecology Letters*, 18(6), 497–503.

- Rokas, A., Atkinson, R. J., Brown, G. S., West, S. A., & Stone, G. N. (2001). Understanding patterns of genetic diversity in the oak gallwasp *Biorhiza pallida*: demographic history or a *Wolbachia* selective sweep? *Heredity*, 87(3), 294–304. <https://doi.org/10.1046/j.1365-2540.2001.00872.x>.
- Rokas, A., Atkinson, R. J., Webster, L., Csoka, G., & Stone, G. N. (2003). Out of Anatolia: longitudinal gradients in genetic diversity support an eastern origin for a circum-Mediterranean oak gallwasp *Andricus quercustozae*. *Molecular Ecology*, 12(8), 2153–2174.
- Satler, J. D., & Carstens, B. C. (2017). Do ecological communities disperse across biogeographic barriers as a unit? *Molecular Ecology*, 26(13), 3533–3545. <https://doi.org/10.1111/mec.14137>.
- Schrider, D. R., Shanku, A. G., & Kern, A. D. (2016). Effects of linked selective sweeps on demographic inference and model selection. *Genetics*, 204(3), 1207–1223.
- Smith, B. T., McCormack, J. E., Cuervo, A. M., Hickerson, M. J., Aleixo, A., Cadena, C. D., Pérez-Emán, J., Burney, C. W., Xie, X., Harvey, M. G., Faircloth, B. C., Glenn, T. C., Derryberry, E. P., Prejean, J., Fields, S., & Brumfield, R. T. (2014). The drivers of tropical speciation. *Nature*, 515(7527), 406–409.
- Smith, J. M., & Haigh, J. (1974). The hitch-hiking effect of a favourable gene. *Genetics Research Cambridge*, 23, 23–35.
- Speidel, L., Forest, M., Shi, S., & Myers, S. R. (2019). A method for genome-wide genealogy estimation for thousands of samples. *Nature Genetics*, 51(9), 1321–1329.
- Stanke, M., & Morgenstern, B. (2005). AUGUSTUS: a web server for gene prediction in eukaryotes that allows user-defined constraints. *Nucleic Acids Research*, 33(Web Server issue), W465–W467.
- Stone, G. N., Challis, R. J., Atkinson, R. J., Csoka, G., Hayward, A., Melika, G., Mutun, S., Preuss, S., Rokas, A., Sadeghi, E., & Schönrogge, K. (2007). The phylogeographical clade trade: tracing the impact of human-mediated dispersal on the colonization of northern Europe by the oak gallwasp *Andricus kollari*. *Molecular Ecology*, 16(13), 2768–2781.
- Stone, G., Lohse, K., Nicholls, J., Fuentes-Utrilla, P., Sinclair, F., Schönrogge, K., Csoka, G., Melika, G., Nieves-Aldrey, J.-L., Pujade-Villar, J., Tavakoli, M., Askew, R., & Hickerson, M. (2012). Reconstructing community assembly in time and space reveals enemy escape in a western palearctic insect community. *Current Biology*, 22(6), 532–537.
- Stone, G. N., Schönrogge, K., Atkinson, R. J., Bellido, D., & Pujade-Villar, J. (2002). The population biology of oak gall wasps (Hymenoptera : Cynipidae). *Annual Review of Entomology*, 47(1), 633–668.
- Stone, G. N., White, S. C., Csoka, G., Melika, G., Mutun, S., Péntes, Z., Sadeghi, S. E., Schönrogge, K., Tavakoli, M., & Nicholls, J. A. (2017). Tournament ABC analysis of the western Palearctic population history of an oak gall wasp, *Synergus umbraculus*. *Molecular Ecology*, 26(23), 6685–6703.
- Tison, J.-L., Edmark, V. N., Sandoval-Castellanos, E., Van Dyck, H., Tammaru, T., Välimäki, P., Dalén, L., & Gotthard, K. (2014). Signature of post-glacial expansion and genetic structure at the northern range limit of the speckled wood butterfly. *Biological Journal of the Linnean Society*, 113(1), 136–148.
- Van der Auwera, G. A., Carneiro, M. O., Hartl, C., Poplin, R., del Angel, G., Levy-Moonshine, A., Jordan, T., Shakir, K., Roazen, D., Thibault, J., Banks, E., Garimella, K. V., Althuler, D., Gabriel, S., & DePristo, M. A. (2013). From FastQ data to high-confidence variant calls: The genome analysis toolkit best practices pipeline. *Current Protocols in Bioinformatics*, 43(1), 11.10.1–11.10.33.
- Vitales, D., García-Fernández, A., Garnatje, T., Pellicer, J., & Vallès, J. (2016). Phylogeographic insights of the lowland species *Cheilophus sempervirens* in the southwestern Iberian peninsula. *Journal of Systematics and Evolution*, 54(1), 65–74.
- Wade, M. J. (2007). The co-evolutionary genetics of ecological communities. *Nature Reviews Genetics*, 8(3), 185–195.
- Wakeley, J. (2008). *Coalescent theory: An introduction*. W. H. Freeman.
- Wall, J. D. (2003). Estimating ancestral population sizes and divergence times. *Genetics*, 163(1), 395–404.
- Wan, T., Oaks, J. R., Jiang, X.-L., Huang, H., & Knowles, L. L. (2021). Differences in quaternary co-divergence reveals community-wide diversification in the mountains of southwest China varied among species. *Proceedings of the Royal Society B: Biological Sciences*, 288(1942), 20202567.
- Weinersmith, K. L., Liu, S. M., Forbes, A. A., & Egan, S. P. (2017). Tales from the crypt: a parasitoid manipulates the behaviour of its parasite host. *Proceedings of the Royal Society B: Biological Sciences*, 284(1847), 20162365.
- Wolfram Research, I. (2016). *Mathematica*. Wolfram Research.
- Xue, A. T., & Hickerson, M. J. (2020). Comparative phylogeographic inference with genome-wide data from aggregated population pairs. *Evolution*, 74(5), 808–830.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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